RESEARCH ARTICLE

Effect of 3% saline and furosemide on biomarkers of kidney injury and renal tubular function and GFR in healthy subjects – a randomized controlled trial

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Abstract

Background: Chloride is speculated to have nephrotoxic properties. In healthy subjects we tested the hypothesis that acute chloride loading with 3% saline would induce kidney injury, which could be prevented with the loop-diuretic furosemide.

Methods: The study was designed as a randomized, placebo-controlled, crossover study. Subjects were given 3% saline accompanied by either placebo or furosemide. Before, during and after infusion of 3% saline we measured glomerular filtration rate (GFR), fractional excretion of sodium (FE_{Na}), urinary chloride excretion (u-Cl), urinary excretions of aquaporin-2 (u-AQP2) and epithelial sodium channels (u-ENaC_y), neutrophil gelatinase-associated lipocalin (u-NGAL) and kidney injury molecule-1 (u-KIM-1) as marker of kidney injury and vasoactive hormones: renin (PRC), angiotensin II (p-AngII), aldosterone (p-Aldo) and arginine vasopressin (p-AVP). Four days prior to each of the two examinations subjects were given a standardized fluid and diet intake.

Results: After 3% saline infusion u-NGAL and KIM-1 excretion increased slightly (u-NGAL: 17 ± 24 during placebo vs. -7 ± 23 ng/min during furosemide, p = 0.039, u-KIM-1: 0.21 ± 0.23 vs -0.06 ± 0.14 ng/ml, p < 0.001). The increase in u-NGAL was absent when furosemide was given simultaneously, and the responses in u-NGAL were not significantly different from placebo control. Furosemide changed responses in u-KIM-1 where a delayed increase was observed. GFR was increased by 3% saline but decreased when furosemide accompanied the infusion. U-Na, FE_{Na}, u-Cl, and u-osmolality increased in response to saline, and the increase was markedly pronounced when furosemide was added. FE_K decreased slightly during 3% saline infusion, but simultaneously furosemide. U-ENaC_Y decreased to the same extent after 3% saline infusion in the two groups. 3% saline significantly reduced PRC, p-AngII and p-Aldo, and responses were attenuated by furosemide. p-AVP was increased by 3% saline, with a larger increase during furosemide.

Conclusion: This study shows minor increases in markers of kidney injury after 3% saline infusion Furosemide abolished the increase in NGAL and postponed the increase in u-KIM-1. The clinical importance of these findings needs further investigation.

Trial registration: (EU Clinical trials register number: 2015–002585-23, registered on 5th November 2015)

Keywords: Hypertonic saline, 3% saline, Hyperchloremic acidosis, NGAL, KIM-1, Fractional excretion of sodium

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Background

In critically ill patients and patients undergoing surgery intravenous fluid treatment is an important part of maintaining cardiovascular homeostasis. Crystalloids and colloids are widely used as fluid resuscitation [1-3]. Crystalloids differ in electrolyte composition. Crystalloids with a high content of sodium and chloride such as isotonic saline induce hyperchloremic metabolic acidosis compared to solutions with a lower sodium and chloride content, particularly when administered in higher doses [4–7]. Chloride and hyperchloremic acidosis may impair renal blood flow and induce kidney injury [4, 8–11]. This was first demonstrated in animal experiments, where high chloride concentration during renal perfusion was associated with increased renal vasoconstriction and reductions in renal blood flow and glomerular filtration rate [9, 11]. In healthy subjects isotonic saline compared to infusion with fluids with lower sodium and chloride contents decreased renal blood flow (RBF). [10] In patients submitted to an emergency department, infusion of low chloride containing solutions was associated with a lesser degree of AKI compared to fluid solutions with a higher chloride content [4, 7]. In the clinical setting however the importance of dyschloremia and infusion of high chloride containing solutions is still under much debate [12–14].

In daily practice plasma creatinine is used to estimate renal function. In case of acute kidney injury (AKI) changes in creatinine are seen within days. Novel biomarkers such as neutrophil gelatinase-associated (NGAL) and kidney injury molecule-1 (KIM-1) are within hours able to detect kidney injury and predict the risk of renal replacement therapy and chronic kidney disease (CKD). [15–19] KIM-1 is produced in the proximal tubulus and NGAL in the distal tubulus, and can both be detected in the urine during very little kidney injury [19].

We therefore hypothesized that a large load of chloride given as 3% saline will induce hyperchloremic acidosis and a subsequent kidney injury, which can be detected by measuring glomerular filtration rate (GFR), renal tubular function, and biomarkers of AKI in the urine. In addition, we hypothesized that furosemide impairs kidney damage induced by 3% saline.

We investigated these hypotheses in a study designed as a randomized, placebo-controlled, crossover study were subjects were given 3% saline accompanied by either placebo or furosemide on two separate occasions, where renal function, urinary excretion of biomarkers of kidney injury, and plasma concentrations of vasoactive hormones were measured.

Methods

Subjects

Screening examination included physical examination, medical history, ECG, office BP, clinical biochemistry and urinary albumin analysis.

Inclusion criteria

Healthy women and men, age 18–40 years, BMI 18.5– 30.0 kg/m². Exclusion criteria: History with or clinical signs of diseases in the central nervous system, lungs, thyroid gland, heart, liver or kidneys, diabetes mellitus or malignancies. Clinical important deviations in screening blood or urinary samples, office blood pressure > 140 mmHg systolic and/or > 90 mmHg diastolic, nursing or pregnancy, alcohol or drug abuse, smoking, allergy or intolerance towards furosemide or unwillingness to participate. Withdrawal criteria: Symptoms of hypotension or office BP repeatedly below 50 mmHg diastolic and/or 90 mmHg systolic. Development of exclusion criteria.

Design

The study was a placebo-controlled, randomized, single-blinded, crossover trial. After inclusion subjects were allocated to treatment via computer-generated randomization in blocks of six. Consequently, the subjects received glucose (placebo) or furosemide in a random order on 2 separate examination days. Awashout period of at least 14 days was required between examinations.

Study drugs

Hypertonic saline (3% NaCl, Skanderborg Pharmacy, Skanderborg, Denmark) was given intravenously as continuous infusion (7 ml/kg/hour) for 60 min. Furosemide (Furix, 2 ml of 10 mg/ml, Takeda Pharma, Osaka, Japan) and isotonic glucose (2 ml 50 g glucosemonohydrate/l, Fresenius, Bad Homburg vor der Höhe, Germany) were identical in appearance to the study subjects. Furosemide was administered at a dose of 20 mg (2 ml).

Effect variables

The primary effect variable was u-NGAL. Secondary effect variables were free water clearance (C_{H2O}), GFR, (fractional excretion of sodium) FE_{Na}, (fractional excretion of potassium) FE_K, u-albumin, u-KIM-1, urinary excretions of aquaporin-2 (u-AQP2) and epithelial sodium channels (u-ENaC_{γ}), plasma and urinary osmolality, plasma concentration of renin (PRC), angiotensin II (p-AngII), aldosterone (p-Aldo) and vasopressin (p-AVP), brachial systolic and diastolic blood pressure (DBP, SBP) and heart rate (HR).

Recruitment

Subjects were consecutively recruited by announcements in local newspapers in community Holstebro, Denmark. After written and oral information that included safety concerns by 3% saline and furosemide infusion, a written consent was obtained. A clinical history was gives and examination was performed, blood and urine samples were drawn and ECG was performed to ensure that the subject fulfilled the inclusion criteria and did not meet exclusion criteria.

Number of subjects

With a significance level of 5% and a power of 80% a total of 23 subjects were needed to detect an 85 ng difference in u-NGAL (SD 144 ng). During examination incomplete voiding was expected in some participants. Hence we estimated that 27 subjects should complete the study.

Experimental procedure

Examinations were carried out after 4 days of standardized diet and fluid intake [20–23]. The diet comprised three main meals and three minor meals. Subjects were instructed to eat variedly from the diet until satiated. The diet contained 11,000 kJ/day, was composed of 55% carbohydrates, 15% protein and 30% fat, and ensured a sodium intake of 150 mmol daily. Fluid intake was 2.5 L per day. Two cups of tea or coffee were allowed daily. No alcohol consumption was allowed.

Collection of 24-h urine samples were performed before each examination. The 24-h urine collection was analyzed for sodium, potassium, chloride, osmolality, creatinine, albumin, AQP2, ENaC_y, NGAL and KIM-1.

After an overnight fast, subjects arrived at 8 AM. Two indwelling catheters for blood sampling and administration of 3% saline and furosemide or glucose (placebo) and ⁵¹Cr-EDTA, were placed in cubital veins, one in each arm. Every 30 min after arrival, participants received an oral water load of 175 ml. Subjects were kept in a supine position in a quiet, temperature-controlled room (22–25 °C). Only exception from the supine position was that when urine was collected by voiding in sitting or standing position. At 10.30 AM 3% saline was given as a continuous infusion for 60 min (7 ml/kg/hour) [20]. Furosemide (20 mg in 2 ml) or glucose (2 ml) was given at 10.30 AM according to randomization.

Blood and urine samples were collected every 30 min from 9:30 AM to 2.30 PM, except for the period between 11 and 12 AM and 1.30 PM to 2.30 PM, blood and urine only was collected once. Urine collections were analyzed for potassium, sodium, chloride, ⁵¹Cr-EDTA, creatinine, osmolality, AQP2, ENaC_{γ}, NGAL and KIM-1. The first three clearance periods from 9:00 AM to 10.30 AM were defined as baseline period.

Blood samples were drawn at 10.30 AM (baseline), 11.30 AM (after 60 min of 3% saline infusion), and at 1 PM (90 min after termination of infusion) for determination of p-AVP, p-Aldo, PRC and p-Ang II.

Urinary spot samples were collected 1 and 3–5 days after the examination days. These samples were analyzed for potassium, sodium, chloride, creatinine, osmolality, NGAL, KIM-1, AQP2 and ENaC_y.

Blood pressure measurements

Office BP used at inclusion was measured using the semiautomatic, oscillometric device, Omron 705IT (Omron Matsusaka CO. Ltd., Matsusaka City, Japan). BP during examination were measured using the automatic oscillometric device, Mobil-O-Graph PWA (Medidyne A/S, Nærum, Denmark). BP was measured as double measurements every 30 min from 9:30 AM to 2.30 PM, except for the period between 11 and 12 AM and 1.30 PM to 2.30 PM, where blood pressure only was measured once. The first 4 measurements were defined as baseline.

Biochemical analyses

Urine samples were stored frozen at -20 °C until analyzed. U-AQP2 and u-ENaC_Y were measured by using radioimmunoassays (RIA) as previously described [20– 25]. Antibodies were raised in rabbits to synthetic peptides for AQP2 and ENaC_Y as previously described [20, 23, 26, 27]. The antibodies against AQP2 and ENaC_Y was a gift from Professor Robert Fenton and Professor Søren Nielsen and, The Water and Salt Research Center, Institute of Anatomy, Aarhus University, Denmark.

Blood samples collected for measurements of vasoactive hormones were centrifuged and plasma was separated, and kept frozen until assayed as previously described [26]. AVP and Ang II were extracted from plasma and then determined by RIA [26, 28, 29]. PRC was determined by immunoradiometric assay as previously described [26]. Aldo was determined by RIA as previously described [26].

A commercial enzyme-linked immunosorbent assay (ELISA) from Bioporto (Hellerup, Denmark) was used to determine the u-NGAL [30]. Minimal detection level was 1.4 pg/ml. Variations were interassay max 8% and intraassay max 14%.U -KIM-1 was determined with a commercial enzyme-linked ELISA-kit (Quantijine ELISA) from R&D Systems. Minimal detection level was 3.0 pg/ml. Variations were interassay max 7.8% and intraassay max 4.4% All samples were analyzed with kits from the same batch.

GFR was estimated using constant infusion clearance technique with ⁵¹Cr-EDTA as reference marker. A GFR variation og 15% variation or more between the three baseline periods led to the exclusion of clearance related analysis [20, 22].

Urine and plasma concentration of potassium, sodium, chloride, creatinine, albumin and were determined at the Department of Clinical Biochemistry by routine methods.

Calculations

 $C_{\rm H2O}$ was calculated with the formula $C_{\rm H2O}$ = UO – $C_{\rm osm}$, where $C_{\rm osm}$ is osmolar clearance and UO is urinary output.

 FE_{Na} and FE_{K} were calculated using to the formula FE_{X} = $(X_{u} \ ^{*} \ V \ / \ X_{p})/GFR.$ V is urine flow in ml/min and X_{u} and X_{p} are urine and plasma concentrations of X. In 24-h urine creatinine clearance was used as an estimation of GFR.

Statistics

Data are presented as means \pm standard deviations (SD), when normality was present. If normality was not presenta data are presented as medians with 25 and 75% percentiles in brackets. A paired comparison between and within groups was performed with paired t-test or Wilcoxon signed rank test. To test for deviation during experimental procedure a general linear model for repeated measures (GLM) was performed. If data did not show normality they were logarithmic transformed before GLM. Friedman's test was used to test for deviations within treatment of vasoactive hormones. Correlations were performed with Pearson correlation. Statistical significance was defined as p < 0.05. Statistical analyses were performed using PASW version 20.0.0 (SPSS Inc.; Chicago, IL, USA).

Results

Demographics

Thirty-two subjects were screened for participation in the study. Exclusion was made for eight subjects due to anaemia (1) and withdrawal of consent (7). Thus, 24 patients were included and completed the trial. The 24 subjects (12 females, 12 males), had a mean BMI 23.7 \pm 2.8 kg/m², age 23 \pm 5 years, office BP 123/70 \pm 9/8 mmHg, p-creatinine 72 \pm 13 µmol/L, urine albumin 8 (1;10) mg/L, p-hemoglobin 8.8 \pm 0.8 mmol/L.

GFR and tubular function during baseline conditions

In 24-h urinary collection made prior to the two examinations sodium (u-Na,) FE_{Na} and chloride (u-Cl) excretion rate was slightly but significantly lower prior to furosemide compared to placebo (Table 1). Urine output, C_{H2O} , urinary excretions of potassium and creatinine, FE_{K} , creatinine clearance, UAER, U-AQP-2, u-ENaC_{γ}, u-NGAL and u-KIM-1 were not significantly different between treatments (Table 1).

Similar results were found at baseline during examinations. At baseline during examinations urine output, C_{H2O} , urinary excretions of potassium, FE_K, GFR, UAER, U-AQP-2, u-ENaC_Y, u-NGAL and u-KIM-1 were similar between treatment arms (Tables 3, 4 and 5).

Bodyweight

At baseline body-weight was similar on the two examinations (74.4 ± 11.9 kg before placebo vs 73.8 ± 11.3 kg before furosemide, p = 0.860). After 3% saline and placebo body-weight increased to 74.9 ± 11.9 kg (p < 0.001) and when furosemide was given simultaneously body-weight decreased to $72.8 \pm 11.3 \text{ kg} (p < 0.001)$. The two responses in bodyweight were significantly different between treatments $(0.5 \pm 0.4 \text{ kg vs.} -1.0 \pm 0.5 \text{ kg}, p < 0.001)$.

Plasma electrolytes

Plasma-Na, p-Cl, p-K, p-osmolality and p-total carbon dioxide were similar at baseline. (Table 2). Plasma-Na, p-Cl and p-osmolality increased after 3% saline. Furosemide did not change the response to 3% saline regarding p-Na, but the increase after 3% saline was less pronounced for p-Cl and increased for p-osmolality when furosemide was given (p < 0.001).

P-K decreased in response to 3% saline and the decrease was more pronounced when furosemide was given. P-total carbon dioxide decreased in response to 3% saline but was unchanged in during furosemide. Responses in p-K and p-total carbon dioxide were significantly different after furosemide compared to placebo (Table 2). There was no correlation between the responses to 3% saline between p-Cl and p-total carbondioxide (p = 0.486) and p-K and p-total carbon dioxide (p = 0.895).

GFR and tubular function during 3% saline and furosemide

Table 3 shows the effect of 3% saline and furosemide induced changes in GFR, urine output (UO), $C_{\rm H2O}$, u-Na (excretion rate), $\rm FE_{Na}$, $\rm FE_{K}$ and u-osmolality. Using a general linear model, expected different response patterns during both 3% saline and furosemide compared to 3% saline alone was demonstrated. UO decreased after 3% saline but increased markedly when saline infusion was accompanied by furosemide. In contrast GFR increased after 3% saline and decreased after furosemide treatment. $C_{\rm H2O}$ decreased after 3% saline but the decrease was initially less pronounced when furosemide was given.

U-Na, FE_{Na} , u-Cl, and u-osmolality increased in response to saline and placebo and the increase was sustained throughout the examination. The increase was markedly pronounced when furosemide was given instead of placebo. After furosemide, the increases in u-Na, FE_{Na} , u-Cl, and u-osmolality were however not sustained during the examination and decreased towards baseline values although it was still significantly higher in the last clearance period compared to baseline.

 FE_K decreased slightly during 3% saline infusion. After infusion FE_K returned to baseline level. As expected furosemide increased FE_K with a substantial rapid response that declined during the clearance periods. The increase was maintained until the last two clearance periods.

Markers of kidney injury

U-NGAL and u-KIM-1 excretion rates were similar between examination days at baseline (Fig. 1). U-NGAL

Table 1 24-h urine collection prior to two examinations in a randomized, cross-over study of 24 healthy subjects

	Placebo	Furosemide	<i>P</i> -value
Urine output (mL/minute)	1.84 ± 0.36	1.73 ± 0.39	0.242
C _{H2O} (mL/minute)	-0.23 ± 0,61	-0.15 ± 0.38	0.436
U-creatinine (mmol/24 h)	15.5 ± 4.1	15.2 ± 4.0	0.917
Creatinine clearance (mmol/mL pr. m ²)	134 ± 24	130 ± 19	0.753
U-Na (mmol/24 h)	124 ± 37	100 ± 28	0.017
FE _{Na} (%)	0.62 ± 0.19	0.57 ± 0.9	0.016
U-CI (mmol/24 h)	128 ± 31	108 ± 27	0.052
U-K (mmol/24 h)	62 ± 14	62 ± 23	0.601
FE _K (%)	10.8 ± 2.3	11.1 ± 4.4	0.438
UAER (mg/24 h)	7 (4;10)	7 (5;9)	0.440
U-AQP-2/min (ng/minute)	0.81 ± 0.31	0.77 ± 0.20	0.562
U-AQP-2/creatinine (ng/mmol)	75 ± 15	76 ± 22	0.826
U-ENaC _y / min (ng/minute)	0.79 ± 0.30	0.71 ± 0.25	0.430
U-ENaC $_{\gamma}$ /creatinine (ng/mmol)	77 ± 30	70 ± 24	0.327
U-NGAL / min (ng/min)	16 (7;43)	15 (7;27)	0.063
U-NGAL /creatinine (ng/mmol)	1401 (649;4777)	1409 (524;3433)	0.109
U-KIM-1 / min (ng/min)	0.41 ± 0.21	0.41 ± 0.20	0.580
U-KIM-1 /creatinine (ng/mmol)	39 ± 23	40 ± 19	0.831

Urine output, C_{H2O} free water clearance, *U-Na* urine excretion of sodium, and *U-K* potassium, FE_{Na} fractional excretion of sodium, and FE_K potassium, creatinine clearance, *UAER* urinary excretions rates of albumin, *u-AQP-2/min* aquaporin-2, *u-ENaC_y/min* γ -fraction of the epithelial sodium channel, *u-NGAL/min* neutrophil gelatinase-associated lipocalin and *u-KIM-1/min* kidney injury molecule-1 and in relation to creatinine (u-AQP-2/creatinine, u-ENaC_y/creatinine, u-NGAL/creatinine, u-KIM-1/creatinine. Urine were collected from 07.00 am on the day before the day of examination day to 07.00 am on the day of examination. Data are shown as means ± SD in brackets or medians with 25 and 75 percentiles in brackets. Statistics are performed with paired t-test or Wilcoxon signed rank test

increased slightly after 3% saline and placebo with a significant increase from baseline in the clearance period just after saline infusion was stopped (Fig. 2a, p = 0.034). In this period where the highest level of u-NGAL during placebo was observed, the response from baseline was significantly different from the response in furosemide group (Fig. 2a). However, when the entire examination was examined there was no difference in response between placebo and furosemide (p = 0.104 using GLM).

U-KIM-1 increased after 3% saline and placebo in the two clearance periods (150–210 min) following 3% saline infusion (Fig. 2b, p < 0.05). In the period from 150 to 180 min u-KIM-1 levels were highest and there was a significant difference in response from baseline compared with furosemide (Fig. 2b).

During furosemide no immediately increase in u-KIM-1 was observed, but u-KIM-1 increased in the last two clearance periods compared to placebo for both periods (Period 210–240 min: -0.15 ± 0.18 in placebo vs. 0.21 ± 0.20 in furosemide, p < 0.001; Period 240–300 min: -0.13 ± 0.12 vs. 0.14 ± 0.14 , p < 0.001. Using a GLM the response in u-Kim-1 after 3% saline was significantly changed by furosemide (p < 0.001).

When u-NGAL and u-KIM-1 were adjusted for urinary creatinine excretion similar result as excretion rate were observed (data not shown).

ENaC, AQP2 and UAER

Table 4 shows the effect of 3% saline and furosemide induced changes in u-AQP2, u-ENaC_Y and and UAER. U-AQP2 increased after 3% saline, and the increase was present after saline infusion was stopped. The response in u-AQP2 to 3% saline was changed by furosemide. U-AQP was markedly increased after furosemide during saline infusion compared to placebo. The following periods u-AQP2 decreased to baseline levels. U-ENaC_Y decreased to the same extent after 3% saline infusion in the two groups. UAER was not changed by 3% saline or the combination with furosemide.

Vasoactive hormones in plasma

Plasma-AVP, PRC, p-Ang II and p-Aldo were similar at baseline (Table 5). 3% saline significantly increased AVP and the increased was more pronounced when furosemide was given with 3% saline. 3% saline significantly decreased PRC, p-AngII and p-Aldo. The responses in PRC, p-Ang II and p-Aldo to 3% saline were all significantly attenuated by furosemide.

Blood pressure (BP)

Hemodynamic variables are shown in Table 6. Systolic BP (SBP) was not altered by 3% saline, but diastolic BP (DBP) decreased. Furosemide changed the responses.

Period	Baseline (90 min)	After 60 min hypertonic saline infusion (150 min)	90 min post hypertonic saline infusion (240 min)	P-value (difference in response)
p-Na (mmol/L)				
Placebo	140 ± 2	$144 \pm 2^{*}$	$141 \pm 2^{*}$	0.073
Furosemide	139±2	$144 \pm 2^{*}$	$141 \pm 2^{*}$	
p-K (mmol/L)				
Placebo	3.8 ± 0.2	$3.7 \pm 0.2^{*}$	$4.0 \pm 0.2^{*}$	0.001
Furosemide	3.7 ± 0.2	$3.5 \pm 0.2^{*, +}$	$3.8\pm0.2^{\dagger}$	
p-Cl(mmol/L)				
Placebo	105 ± 2	$111 \pm 2^{*}$	$107 \pm 2^{*}$	< 0.001
Furosemide	104 ± 2	$108 \pm 2^{*, +}$	$104 \pm 2^{*, +}$	
p-Osmolality (mm	nol/L)			
Placebo	282 ± 4	$289 \pm 3^{*}$	$286 \pm 4^{*}$	0.034
Furosemide	282 ± 3	291 ± 3 ^{*, †}	$286 \pm 3^{*}$	
p-total carbondio:	xide (mmol/L)			
Placebo	27 ± 2	$25 \pm 2^{*}$	$25 \pm 2^{*}$	< 0.001
Furosemide	26 ± 2	$26 \pm 2^{\dagger}$	$27 \pm 2^{\dagger}$	

Table 2 Effect of hypertonic saline and furosemide on plasma concentrations of electrolytes in a randomized, cross-over study of 24 healthy subjects

p-*Na* Plasma concentrations of sodium, *p*-*K* potassium, *p*-*Cl* chloride and total carbondioxide and plasma osmolality were measured every 30 min during examination. Data show are values before hypertonic saline infusion, after 60 min of saline infusion, and 90 min after cessation of saline infusion on the examination day. Data are shown as medians with 25 and 75 percentiles in brackets. P-value represents probability of difference in response to saline (response from baseline to saline infusion) between treatments. To test difference in response to saline between treatments a students t-test was used. Wilcoxon signed rank test was performed to test differences from baseline, * = p < 0.05, and from Placebo, $^+ = p < 0.05$

When furosemide was given along with 3% saline SBP decreased. DBP also decreased but decreased but the response seemed delayed compared to placebo.

Urinary spot samples day 1 and day 3–5 post examination

The results from urinary spot samples are shown in Table 7. The urinary spot sample performed 2 days after examination showed a decreased sodium concentration (u-Na) and increased potassium (u-K), creatinine and albumin concentration after furosemide compared to placebo (Table 7). Urine osmolality was increased after furosemide. Urinary chloride concentration (u-Cl), u-NGAL, u-KIM-1, u-AQP2 and u-ENaC_{γ} were not significally different.

The urinary spot sample performed 3-5 days after examination, revealed no difference between the furosemide and placebo treatment for any of the variables in Table 7.

Discussion

The main findings in this study was small increases in u-NGAL and U-KIM-1 after 3% saline. The increase in u-NGAL after 3% saline was abolished by furosemide. The response in u-KIM-1 was changed after furosemide, where the increase in u-KIM-1 after 3% saline was delayed to the last clearance periods. In addition, when furosemide was given along with 3% saline the increased p-Cl was attenuated and the decrease in p-total carbon dioxide was abolished. Although the increases in u-NGAL and u-KIM-1 after 3% saline were small, the increases may support the hypothesis that sodium-chloride solutions are nephrotoxic, but this study does not show convincing evidence for nephroprotective properties of furosemide.

Chloride induced metabolic acidosis after 0.9% saline (isotonic) has been reported previously [4, 6, 8-10, 31, 32]. The hyperchloremic acidosis is at least partly explained by intracellular displacement of the anion bicarbonate by chloride to reduce the anion gap in case of hyperchloremia [33]. A similar finding is also reported after hypertonic saline in healthy subjects where 3% saline increased plasma chloride and caused a respiratory compensated metabolic acidosis [34]. These findings were confirmed in our study were 3% saline infusion increased plasma chloride and evidence of acidosis was suggested by the reduced p-total carbondioxide. Total carbon dioxide is generally a good marker of serum bicarbonate due to the fact that bicarbonate comprises about 95% of total carbondioxide [6]. It is possible that the changes in total carbondioxide were due to changes in other forms of carbondioxide such as dissolved CO₂ or carbonic acid, but most likely the changes are caused by changes in plasma bicarbonate. Furosemide attenuated the increase in plasma chloride and abolished the decrease in total carbondioxide after 3% saline

Period	Baseline	Hypertonic saline infusion	Post hypertoni				
	0–90 min	90–150 min	150–180 min	180–210 min	210–240 min	240–300 min	P (GLM within)
GFR (⁵¹ Cr-EDTA clear	ance)						
Placebo	104 ± 14	102 ± 15	107 ± 15	$110 \pm 15^{*}$	$111 \pm 23^{*}$	$112 \pm 15^{*}$	0.001
Furosemide	104 ± 12	103 ± 13	103 ± 18	$93 \pm 14^{*}$	$93 \pm 14^{*}$	98±12	
P (GLM between)	0.089						
Urine output (mL/mi	in)						
Placebo	9.8 ± 1.5	$3.5 \pm 1.6^{*}$	$2.7 \pm 1.3^{*}$	$2.7 \pm 0.8^{*}$	$3.3 \pm 1.4^{*}$	4.7 ± 2.2 [*]	< 0.001
Furosemide	9.1 ± 2.2	23.1 ± 2.6 [*]	10.1 ± 3.1	$4.3 \pm 1.8^{*}$	$2.4 \pm 0.9^{*}$	$2.0 \pm 1.2^{*}$	
P (GLM between)							
			< 0.001				
C _{H2O} (ml/min)		*	×	×	×	*	
Placebo	6.6 ± 1.3	-0.4 ± 1.4	$-2.2 \pm 1.2^{\circ}$	-2.4 ± 1.0"	-1.8 ± 1.6"	0.0 ± 2.1	0.001
Furosemide	6.1 ± 2.0	1.1 ± 1.1	-2.0 ± 0.7	-1.8 ± 0.5	-1.5 ± 0.5	-1.1 ± 0.6	
P (GLM between)	0.597						
U-Na (µmol/min)			v	v	v		
Placebo	200 ± 94	$361 \pm 146^*$	501 ± 234*	531 ± 182*	511 ± 161*	466 ± 106*	< 0.001
Furosemide	162 ± 78	$2865 \pm 342^*$	1515 ± 3.81 [*]	659 ± 274 [*]	377 ± 158 [*]	273 ± 141*	
P (GLM between)	< 0.001						
FE _{Na} (%)							
Placebo	1.38 ± 0.63	$2.46 \pm 0.85^{*}$	$3.28 \pm 1.49^{*}$	$3.36 \pm 0.89^{*}$	$3.26 \pm 0.81^{*}$	$3.00 \pm 0.69^{*}$	< 0.001
Furosemide	1.13 ± 0.55	19.81 ± 3.11 [*]	10.66 ± 3.81*	5.03 ± 1.97 [*]	3.03 ± 1.59 [*]	2.05 ± 1.31*	
P (GLM between)	< 0.001						
U-Cl (µmol/min)							
Placebo	239 ± 84	$379 \pm 146^{*}$	$537 \pm 261^{*}$	$575\pm201^{*}$	$558 \pm 185^{*}$	$502 \pm 122^{*}$	< 0.001
Furosemide	212 ± 61	$3083 \pm 356^{*}$	$1679 \pm 462^{*}$	$763 \pm 298^{*}$	$441 \pm 177^{*}$	$310 \pm 156^{*}$	
P (GLM between)	< 0.001						
FE _K (%)							
Placebo	21.1 ± 6.2	18.1 ± 7.1*	21.6 ± 17.0	23.1 ± 10.0	22.7 ± 8.0	21.9 ± 7.8	< 0.001
Furosemide	24.0 ± 9.2	64.9 ± 15.2 [*]	44.6 ± 14.7 [*]	34.6 ± 16.6 [*]	27.4 ± 11.4	23.8 ± 10.5	
P (GLM between)	< 0.001						
U-osmolality (µmol//	'min)						
Placebo	899 ± 205	1103 ± 304 [*]	$1416 \pm 592^{*}$	$1485 \pm 403^{*}$	$1446 \pm 351^{*}$	$1351 \pm 260^{*}$	< 0.001
Furosemide	831 ± 124	$6293 \pm 684^{*}$	$3514 \pm 881^{*}$	$1746 \pm 602^{*}$	$1129 \pm 323^{*}$	905 ± 328	
P (GLM between)	< 0.001						

Table 3	B Effect	of hypert	onic salin	e and	furosemide	on (GFR and	tubular	function	in a	randomized,	cross-over	study	of 2	24 he	althy
subjects	5															

GFR Glomerular filtration rate, urine output, C_{H2O} free water clearance, *u*-*Na/min* urinary sodium excretion, *FE_{Na}* fractional excretion of sodium, *u*-*Cl/min* urinary chloride excretion and *FE_k* fractional excretion of potassium, Urine was collected every 30 min in the 90 min baseline period, once after 60 min of hypertonic infusion, and every 30 min 90 min after hypertonic saline infusion and once 150 min after cessation of hypertonic saline infusion. Data from three baseline periods are pooled and shown as one period. Data are presented as means ± SD. Statistics are performed with a general linear model (GLM) or paired t-test. Difference from baseline: * = p < 0.05

Assuming that total carbondioxide is a marker of bicarbonate, furosemide seems to prevent the metabolic acidosis induced by 3% saline. Metabolic alkalosis due to increased renal bicarbonate excretion is a known adverse reaction after furosemide treatment, although the renal mechanisms are not fully understood [35]. We measured two novel markers of kidney injury in the urine, NGAL and KIM-1, that are related to increased risk of renal replacement therapy and CKD in in patients with AKI [15–19]. Both u-NGAL and u-KIM-1 were slightly but significantly increased by 3% saline, suggesting renal injury induced by the hypertonic saline



load. 3% saline increased GFR and decreased UO, which could influence the increase, but the increase was present when excretion was adjusted for urinary volume (flow) and creatinine excretion, so it is unlikely that that changes in GFR and UO are the explanation for the increased u-NGAL and u-KIM-1. Urine composition changed as expected after furosemide, with an increased osmolality and excretion of sodium and chloride, and these changes could have influenced the excretion of u-NGAL and u-KIM-1 without any kidney injury. However, it is unknown if marked changes in tubular electrolyte composition can change the excretion of u-NGAL and u-KIM-1. In spontaneously hypertensive rats high salt intake increased urinary NGAL and KIM-1 indicating that high dietary salt induces kidney injury [36]. High salt in this rat model was accompanied by an increased BP which is also likely to explain the increased urinary excretion of markers in kidney injury rather than salt intake itself. In the present study the salt load seemed to decrease BP rather than increase excluding blood pressure as a mediator of the increase in markers of kidney injury. Chloride and hyperchloremic acidosis



has previously been demonstrated to influence renal hemodynamics by impairing RBF [9–11]. This is in contrast to observations in patients with heart failure where hypertonic saline preserved renal function, but no biomarkers were measured in these patients [37]. It is possible that certain patient groups may benefit from hypertonic saline while other patient groups does not. Patients with heart failure tend to be hypotensive and theoretically a volume expansion with 3% saline may increase blood pressure end subsequently RBF. We did not measure RBF and cannot evaluate changes in RBF. GFR was initially unchanged after 3% saline but increased in the last clearance periods which does not support a lowered RBF after 3% saline.

The loop-diuretic furosemide markedly increased UO and electrolyte excretion which was expected [20, 21, 23, 38, 39]. Furosemide attenuated the 3% saline induced increase in p-Cl and abolished the reduction in total

carbondioxide. Hence furosemide attenuated the metabolic acidosis induced by 3% saline. The increases in u-NGAL and u-KIM-1, which were observed in the clearance periods just after 3% saline infusion, were abolished by furosemide. This might suggest renoprotective properties of furosemide. However, the increase in u-NGAL after 3% saline only just reached statistical significance and may be influenced by the huge increase in diuresis during furosemide, which dilutes the concentration of u-NGAL which increases the uncertainty of measurement. In addition, the increase in u-KIM-1 seems delayed after 3% saline and furosemide compared to placebo and was present in the last to clearance periods rather than the periods immediately after saline infusion. Accordingly, furosemide changed the response in u-KIM-1 where when compared to placebo a delayed increase was observed. It still under debate if furosemide is harmful or protective to the kidneys. Furosemide is

Period	Baseline	Hypertonic saline infusion	Post hypertonic				
	0–90 min	90–150 min	150–180 min	180–210 min	210–240 min	240–300 min	P (GLM within)
U-AQP2 (ng/minute	<u>e)</u>						
Placebo	0.81 (0.66;0.93)	0.85 (0.71;1.05)	1.00 (0.81;1.31)*	1.01 (0.87;1.38)*	1.07 (0.77;1.26)*	0.98 (0.80;1.09)*	< 0.001
Furosemide	0.77 (0.66;0.92)	1.42 (1.18;1.61)*	1.12 (0.90;1.41)*	1.14 (0.88;1.46)*	0.86 (0.72;1.10)*	0.79 (0.72;0.93)	
P (GLM between)	0.553						
U-AQP2 /creatinine	(ng/mmol)						
Placebo	72 (66;84)	84 (74;87)	86 (83;104)*	102 (85;111) [*]	90 (77;99)*	92 (82;98)*	< 0.001
Furosemide	76 (63;83)	140 (104;150)*	100 (91;129)*	121 (84;132)*	97 (81;107)*	88 (64;104)*	
P (GLM between)	0.186						
U-ENaC _y (ng/minute	e)						
Placebo	0.87 (0.71;1.27)	0.73 (0.60;1.19)	0.90 (0.76;1.27)	0.81 (0.70;1.10)	0.75 (0.64;1.13)	0.68 (0.60;1.03)*	0.399
Furosemide	0.92 (0.83;1.26)	0.87 (0.75;1.03)	1.06 (0.63;1.31)	0.83 (0.64;1.12)	0.81 (0.64;1.04)*	0.73 (0.63;0.93)*	
P (GLM between)	0.806						
U-ENaC $_{\gamma}$ /creatinine	(ng/mmol)						
Placebo	80 (72;97)	80 (63;90)	84 (72;98)	79 (63;87)	70 (63;89)	69 (62;81)*	0.884
Furosemide	91 (82;99)	75 (66;131)	81 (70;116)	88 (72;16)	83 (69;107)*	72 (60;97)*	
P (GLM between)	0.487						
UAER (µg/min)							
Placebo	1 (0;5)	3 (3;4)	4 (4;6)	4 (3;6)	4 (3;4)	3 (0;4)	0.129
Furosemide	1 (0;5)	0 (0;9)	0 (0;10)	4 (1;7)	4 (2;6)	3 (2;5)	
P (GLM between)	0.167						

Table 4 Effect of hypertonic saline and furosemide on excretion of proteins from epithelial sodium channels and aquaporin-2 channels in a randomized, cross-over study of 24 healthy subjects

u-AQP2/minute Aquaporin-2 excretion rate, *U*-AQP2/creatinine creatinine adjusted u-AQP2 excretion, *u*-ENaC_y/minute excretion of the γ -fraction of the epithelial sodium channel and *U*-ENaC_y /creatinine creatinine adjusted u-ENAC_y. *UAER* urinary albumin excretion rate. Urine was collected every 30 min in the 90 min baseline period, once after 60 min of hypertonic infusion, and every 30 min 90 min after hypertonic saline infusion and once 150 min after cessation of hypertonic saline infusion. Data from three baseline periods are pooled and shown as one period. Data are shown as medians with 25 and 75 percentiles in brackets. *P*-value represents probability of difference in response to hypertonic saline (response from baseline to hypertonic saline) between treatments Statistics are performed with a general linear model (GLM), or Wilcoxon signed rank test. Data were logarithmic transformed before GLM was performed. Difference from baseline : = p < 0.05

shown to increase oxidative stress in the kidneys. [40] A recent meta-analysis did not find evidence of increased risk of AKI when furosemide was given as bolus injections [41]. In intensive care units furosemide is shown not to influence u-NGAL levels or renal prognosis [42, 43]. Although this study demonstrates some signs of positive protective effects of furosemide, further studies are warranted before conclusions can be drawn whether furosemide have harmful or protective properties after saline infusion.

AQP2 is located in the collecting duct principal cells and when inserted in the apical membrane increases water permeability and reabsorption [44]. AVP stimulates this insertion. Due to an increase in plasma osmolality induced by 3% saline the increases in AVP and subsequent increase in u-AQP2 were expected [20, 23]. The increase in AVP and u-AQP2 was further increased when furosemide was given simultaneously, likely explained by diuresis induced intravascular fluid depletion. Increased AVP and u-AQP2 to furosemide are established, and an additive increase in AVP due to the combined effects of 3% saline and furosemide was expected [21, 38, 39]. Hence 3% saline, furosemide and the combination of the two interventions induce increased water-reabsorption in the collecting ducts.

The 3% saline increased plasma osmolality and intravascular volume, and in concordance with our previous studies decreases in PRC, p-AngII and p-Aldosterone [20, 21, 23, 38, 39]. Furosemide caused a decrease in BP probably explained by a diuresis induced intravascular fluid depletion. Similarly, the decrease in the vasoactive hormones PRC, p-AngII and P-Aldo was attenuated and the increase in p-AVP was exaggerated. We have previously demonstrated that fluid depletion induced by furosemide creates increases in concentrations of PRC, p-AngII and p-Aldo [20, 21, 23, 38, 39]. This compensatory response is confirmed in this study where PRC, p-AngII and p-Aldo also increased after furosemide compared to placebo.

	Baseline (90 min)	After 60 min hypertonic saline infusion (150 min)	90 min post hypertonic saline infusion (210 min)	P-value (difference in response)
p-AVP (ng/L)				
Placebo	0.20 (0.20;0.20)	0.50(0.40;0.70)*	0.20(0.20;0.23)	< 0.001
Furosemide	0.20 (0.18;0.20)	0.90(0.60;1.10)* ^{,†}	0.30(0.20;0.40) ^{*,†}	
PRC (ng/L)				
Placebo	9.0 (5.3;13.0)	7.3 (4.4;10.9) [*] 5.6 (2.9;7.4) [*]		0.001
Furosemide	10.3 (5.8;16.9)	9.3 (7.7;16.2) [†]	8.0 (5.3;19.3) [†]	
p-Angll (ng/L)				
Placebo	12 (8;18)	7 (5;13)*	6 (4;11) *	0.014
Furosemide	16 (9;22)	16 (11;20) [†]	16 (9;24) [†]	
p-Aldo (pmol/L)				
Placebo	240 (200;342)	167 (144;211)*	169 (161;213) [*]	0.001
Furosemide	277 (232;377)	256 (228;328) [†]	262 (201;325) [†]	

Table 5 Effect of hypertonic saline and furosemide on vasoactive hormones in a randomized, cross-over study of 24 healthy subjects

p-*AVP* Plasma concentrations arginine vasopressin, *PRC* renin, *p*-*Angll* angiotensin II and *p*-*Aldo* aldosterone were measured before hypertonic saline infusion, after 60 min of saline infusion, and 90 min after cessation of saline infusion on the examination day. Data are shown as medians with 25 and 75 percentiles in brackets. *P*-value represents probability of difference in response to saline (response from baseline to saline infusion) between treatments. Students t-test was used to test difference in response to saline between treatments. Wilcoxon signed rank test was used to test statistical significant difference from baseline, * = p < 0.05, and from Placebo, $^{+} = p < 0.05$

ENaC regulates sodium transport in the distal tubulus. In animal models changes in renal and plasma osmolality changed ENaC abundance in the collecting duct and ENaC activity [45, 46]. In previous studies small increases in u-ENaC_{γ} were observed in response to 3% saline [20, 23]. Hence we expected increases in u-ENaC_{γ} but in this study u-ENaC_{γ} was not changed by 3% saline. ENaC's activity is regulated by aldosterone [47]. In this study p-Aldo

decreased after 3% saline and was unchanged when furosemide was added, which can explain why u-ENaC_Y was unchanged. In addition, we used a higher infusion rate of 3% saline than used in previous studies resulting in a higher total dose of 3% saline, which could explain difference from previous studies of u-ENaC_Y.

Despite being on an identically standardized diet 4 days prior to each examination there was a small but

 Table 6 Effect of hypertonic saline and furosemide on hemodynamic variables in a randomized, cross-over study of 24 healthy subjects

Period	Baseline	Hypertonic saline infusion	Post hypertoni				
	0–90 min	90–150 min	150–180 min	180–210 min	210–240 min	240–300 min	P (GLM within)
SBP (mmHg)							
Placebo	118 ± 9	118 ± 10	117±9	119 ± 10	117±10	120±9	0.001
Furosemide	117 ± 7	120±13	$114 \pm 6^{*}$	111±8 [*]	113±8 [*]	116±8	
P (GLM between)	0.202						
DBP (mmHg)							
Placebo	68 ± 7	67±7	$64 \pm 8^{*}$	$67 \pm 7^{*}$	$66 \pm 6^{*}$	67±8	0.003
Furosemide	68 ± 6	69±7	67±8	$66 \pm 6^{*}$	$67 \pm 7^{*}$	$66 \pm 6^{*}$	
P (GLM between)	0.740						
HR (beats/min)							
Placebo	62 ± 10	$66 \pm 11^{*}$	63±11	$64 \pm 10^{*}$	62 ± 10	$64 \pm 11^{*}$	0.354
Furosemide	61±8	$65 \pm 10^{*}$	$63 \pm 9^{*}$	$63 \pm 8^{*}$	$64 \pm 10^{*}$	$64 \pm 10^{*}$	
P (GLM between)	0.875						

SBP, DBP Systolic and diastolic blood pressure, *HR* heart rate, *cSBP, cDBP* central systolic and diastolic blood pressure, *AI* augmentation index *VR* vascular resistance. Blood pressure was measured every 30 min in the 90 min baseline period, once after 60 min of hypertonic infusion, and every 30 min 90 min after hypertonic saline infusion and once 150 min after cessation of hypertonic saline infusion. Data from four baseline measurements are pooled and shown as one period. Data are presented as means \pm SD. Statistics are performed with a general linear model (GLM) or paired t-test. Statistically significant difference from baseline: * = p < 0.05

	Spot 1 (day 1 post examination)	Spot 2 (day 3–5 post examination)
U-Na (mmol/L)		
Placebo	79 (41;105)	50 (31;135)
Furosemide	57 (27;98) [†]	62 (32;151) [*]
U-K (mmol/L)		
Placebo	26 (15;37)	27 (14;44)
Furosemide	34 (19;52) [†]	30 (17;58)*
U-CI (mmol/L)		
Placebo	78 (53;126)	63 (40;129)
Furosemide	67 (38;116)	70 (45;186)
U-Creatinine (mmol/L)		
Placebo	4 (3;6)	5 (4;16)
Furosemide	8 (4;13) [†]	5 (3;13)*
U-Osmolality (mmol/L)		
Placebo	392 (200;467)	269 (195;710) [*]
Furosemide	430 (236;663) [†]	358 (214;740) [*]
U-Albumin (mg/L)		
Placebo	2 (1;5)	4 (2;7)
Furosemide	5 (3;6) [†]	4 (2;4)
U-ENaC _y (ng/ml)		
Placebo	0.29 (0.18;0.44)	0.35 (0.19;0.85)
Furosemide	0.57 (0.27;0.92)	0.37 (0.21;0.92)
U-AQP2 (ng/ml)		
Placebo	0.40 (0.19;0.49)	0.37 (0.26;0.81)*
Furosemide	0.68 (0.27;0.95)	0.36 (0.29;1.08)*
U-NGAL (ng/ml)		
Placebo	8.5 (3.8;23.8)	9.5 (2.8;22.3)*
Furosemide	19.5 (4.0;40.5)	14.0 (3.0;25.5)*
U-KIM (ng/ml)		
Placebo	0.13 (0.08;0.17)	0.22 (0.09;0.35)*
Furosemide	0.40 (0.09;0.57)	0.20 (0.08;0.49)*

Table 7 Effect of hypertonic saline and furosemide on urinary electrolytes and proteins in two spot urinary sample after examination in a randomized, cross-over study of 24 healthy subjects

u-Na Urinary concentrations of sodium, *u-K* potassium, *u-Cl* chloride, creatinine, albumin, *u-ENaC_y* γ -fraction of the epithelial sodium channel, *u-AQP2* aquaporin 2, *u-NGAL* neutrophil gelatinase-associated lipocalin and *u-KIM-1* kidney injury, molecule-1. Data are shown as medians with 25 and 75 percentiles in brackets. Wilcoxon signed rank test was used to test statistically significant difference from spot 1, * = *p* < 0.05, and from Placebo, [†] = *p* < 0.05

significantly lower sodium excretion in the 24-h prior to the examination where furosemide was given. All other parameters measured in the 24-h urine were not significantly different between examination days. This difference in sodium excretion may have influenced our results but we think it is unlikely because sodium excretion was similar at baseline on examinations days. The urinary spot samples collected day 1 after examination show furosemide changes in urine osmolality, creatinine, potassium and sodium concentration. These changes were not present in the spot urinary samples day 3–5 after examination. This suggest minimal carry-over effects of furosemide, which is a possibility in this cross-over study design.

There were no differences in markers of kidney injury in the post-experiment spot samples suggesting no long term nephrotoxic or nephroprotective effects of furosemide. The spot samples were collected at a random time between 7 AM and 2 PM and days without standardization of the diet, which could cause a larger variation in urine composition and we are therefore cautious to make definite conclusion based on these spot samples.

3% saline was chosen rather than 0.9% saline because we wanted to limit the confounding effects the volume load given with the saline infusion. Since 0.9% saline is mostly used in daily clinical settings and 3% saline is only used in specific cases, this reduces the generability to daily clinical practice. We chose the dose of 7 ml/kg/hour. This resulted in an average infusion dose of approximately 500 ml which we considered sufficient to give to see nephrotoxic effects of a high chloride load without safety concerns. The effect of different doses could reveal diffrences in urine excreation of renal injury but this needs further investigation.

Conclusions

Furosemide given along with 3% saline attenuated the increase in p-Cl and prevented the decrease in p-total carbondioxide induced by 3% saline. The small increases in u-NGAL after 3% saline were abolished by furosemide. The increase in u-KIM-1 induced by hypertonic saline was delayed by furosemide Although the increases in u-NGAL and u-KIM-1 after 3% saline were small, the increases may support the hypothesis that sodium-chloride solutions are nephrotoxic. The changes i p-Cl, p-total carbon dioxide and u-NGAL suggest renoprotective properties as well, but the response in u-KIM-1 does not support this suggestion. Further investigations are warranted before conclusion can be made.

Abbreviations

AKI: Acute kidney injury; Aldo: Aldosterone; AnglI: Angiotensin II; AQP-2: aquaporin-2; AVP: vasopressin; BP: Blood pressure; C_{H2O}: Free water clearance; CKD: Chronic kidney disease; CI: Chloride; C_{OSM}: Osmolar clearance; DBP: brachial diastolic blood pressure; EDTA: Ethylenediaminetetraacetic acid; ENAC_y : Gamma fraction of epithelial sodium channels; FE_K: Fractional excretion of potassium; FE_{Na}: Fractional excretion of sodium; GFR: Glomerular filtration rate; GLM: General linear model; HR: Heart rate; K: Potassium; KIM-1: kidney injury molecule-1; Na: Sodium; NGAL: Neutrophil gelatinaseassociated lipocalin (u-NGAL); PRC: Plasma concentration of renin; RBF: Renal blood flow; RIA: Radioimmunoassay; SBP: brachial systolic blood pressure; UAER: Urinary albumin excretion rate; UO: Urine output

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Availability of data and materials

Access to data can be given by correspondence to FHM.

Authors' contributions

All authors have consented and contributed to the publication. ANJ, NPE, EBP and JNB designed the project. ANJ and MHV performed the experiments and performed laboratory analysis, FHM performed statistical analysis, FHM, ANJ, MHV, NPE, EBP and JNB wrote and edited the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the Regional Committee on Biomedical Research Ethics (case number: 1–10–72-332-15) and Danish Health and Medicines Authority (EudraCT number: 2015–002585-23). An informed, signed consent was obtained from each subject. The study was carried out

in accordance with the Declaration of Helsinki and was monitored by the Good clinical practice-unit from Aarhus and Aalborg Universities.

Consent for publication

Not apllicable.

Competing interests

All authors declare that they have no competing interests.

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References

- Perner A, Haase N, Guttormsen AB, Tenhunen J, Klemenzson G, Åneman A, et al. Hydroxyethyl starch 130/0.42 versus Ringer's acetate in severe Sepsis. N Engl J Med. 2012;367:124–34. https://doi.org/10.1056/NEJMoa1204242.
- Haase N, Perner A, Hennings LI, Siegemund M, Lauridsen B, Wetterslev M, et al. Hydroxyethyl starch 130/0.38-0.45 versus crystalloid or albumin in patients with sepsis: systematic review with meta-analysis and trial sequential analysis. BMJ. 2013;346:f839 http://www.ncbi.nlm.nih.gov/ pubmed/23418281. Accessed 8 May 2018.
- Perner A, Gordon AC, De Backer D, Dimopoulos G, Russell JA, Lipman J, et al. Sepsis: frontiers in diagnosis, resuscitation and antibiotic therapy. Intensive Care Med. 2016;42:1958–69. https://doi.org/10.1007/s00134-016-4577-z.
- Yunos NM, Bellomo R, Glassford N, Sutcliffe H, Lam Q, Bailey M. Chlorideliberal vs. chloride-restrictive intravenous fluid administration and acute kidney injury: an extended analysis. Intensive Care Med. 2015;41:257–64. https://doi.org/10.1007/s00134-014-3593-0.
- Scheingraber S, Rehm M, Sehmisch C, Finsterer U. Rapid saline infusion produces hyperchloremic acidosis in patients undergoing gynecologic surgery. Anesthesiology. 1999;90:1265–70 http://www.ncbi.nlm.nih.gov/ pubmed/10319771. Accessed 10 May 2018.
- Barker ME. 0.9% saline induced hyperchloremic acidosis. J Trauma Nurs. 2015;22:111–6. https://doi.org/10.1097/JTN.00000000000115.
- Self WH, Semler MW, Wanderer JP, Wang L, Byrne DW, Collins SP, et al. Balanced crystalloids versus saline in noncritically ill adults. N Engl J Med. 2018;378:819–28. https://doi.org/10.1056/NEJMoa1711586.
- Yunos NM, Bellomo R, Taylor DM, Judkins S, Kerr F, Sutcliffe H, et al. Renal effects of an emergency department chloride-restrictive intravenous fluid strategy in patients admitted to hospital for more than 48 hours. Emerg Med Australas. 2017;29:643–9. https://doi.org/10.1111/1742-6723.12821.
- Wilcox CS. Regulation of renal blood flow by plasma chloride. J Clin Invest. 1983;71:726–35 http://www.ncbi.nlm.nih.gov/pubmed/6826732. Accessed 10 May 2018.
- Chowdhury AH, Cox EF, Francis ST, Lobo DN. A randomized, controlled, double-blind crossover study on the effects of 2-L infusions of 0.9% saline and plasma-Lyte[®] 148 on renal blood flow velocity and renal cortical tissue perfusion in healthy volunteers. Ann Surg. 2012;256:18–24. https://doi.org/ 10.1097/SLA.0b013e318256be72.
- Bullivant EM, Wilcox CS, Welch WJ. Intrarenal vasoconstriction during hyperchloremia: role of thromboxane. Am J Phys. 1989;256(1 Pt 2):F152–7. https://doi.org/10.1152/ajprenal.1989.256.1.F152.
- 12. Bandak G, Kashani KB. Chloride in intensive care units: a key electrolyte. F1000Research. 2017;6:1930. https://doi.org/10.12688/f1000research.11401.1.
- Yessayan L, Neyra JA, Canepa-Escaro F, Vasquez-Rios G, Heung M, Yee J, et al. Effect of hyperchloremia on acute kidney injury in critically ill septic patients: a retrospective cohort study. BMC Nephrol. 2017;18:346. https://doi. org/10.1186/s12882-017-0750-z.
- 14. Shao M, Li G, Sarvottam K, Wang S, Thongprayoon C, Dong Y, et al. Dyschloremia is a risk factor for the development of acute kidney injury in

critically ill patients. PLoS One. 2016;11:e0160322. https://doi.org/10.1371/journal.pone.0160322.

- Koyner JL, Vaidya VS, Bennett MR, Ma Q, Worcester E, Akhter SA, et al. Urinary biomarkers in the clinical prognosis and early detection of acute kidney injury. Clin J Am Soc Nephrol. 2010;5:2154–65. https://doi.org/10.2215/CJN.00740110.
- Chen L-X, Koyner JL. Biomarkers in acute kidney injury. Crit Care Clin. 2015; 31:633–48. https://doi.org/10.1016/j.ccc.2015.06.002.
- Beker BM, Corleto MG, Fieiras C, Musso CG. Novel acute kidney injury biomarkers: their characteristics, utility and concerns. Int Urol Nephrol. 2018; 50:705–13. https://doi.org/10.1007/s11255-017-1781-x.
- Klein SJ, Brandtner AK, Lehner GF, Ulmer H, Bagshaw SM, Wiedermann CJ, et al. Biomarkers for prediction of renal replacement therapy in acute kidney injury: a systematic review and meta-analysis. Intensive Care Med. 2018;44:323–36. https://doi.org/10.1007/s00134-018-5126-8.
- Moledina DG, Parikh CR. Phenotyping of acute kidney injury: beyond serum creatinine. Semin Nephrol. 2018;38:3–11. https://doi.org/10.1016/j. semnephrol.2017.09.002.
- Jensen JM, Mose FH, Kulik A-EO, Bech JN, Fenton RA, Pedersen EB. Abnormal urinary excretion of NKCC2 and AQP2 in response to hypertonic saline in chronic kidney disease: an intervention study in patients with chronic kidney disease and healthy controls. BMC Nephrol. 2014;15:101.
- Matthesen SK, Larsen T, Vase H, Lauridsen TG, Jensen JM, Pedersen EB. Effect of Amiloride and spironolactone on renal tubular function and central blood pressure in patients with arterial hypertension during baseline conditions and after furosemide: a double-blinded, randomized, placebocontrolled crossover trial. Clin Exp Hypertens. 2013;35:313–24. https://doi. org/10.3109/10641963.2012.721843.
- Mose FH, Jensen JM, Therwani S, Mortensen J, Hansen AB, Bech JN, et al. Effect of nebivolol on renal nitric oxide availability and tubular function in patients with essential hypertension. Br J Clin Pharmacol. 2015;80:425–35.
- Jensen JM, Mose FH, Bech JN, Nielsen S, Pedersen EB. Effect of volume expansion with hypertonic- and isotonic saline and isotonic glucose on sodium and water transport in the principal cells in the kidney. BMC Nephrol. 2013;14:202. https://doi.org/10.1186/1471-2369-14-202.
- Graffe CC, Bech JN, Pedersen EB. Effect of high and low sodium intake on urinary aquaporin-2 excretion in healthy humans. Am J Physiol Renal Physiol. 2012;302:F264–75. https://doi.org/10.1152/ajprenal.00442.2010.
- Pedersen RS, Bentzen H, Bech JN, Pedersen EB. Effect of water deprivation and hypertonic saline infusion on urinary AQP2 excretion in healthy humans. Am J Physiol Renal Physiol. 2001;280:F860–7. https://doi.org/10. 1152/ajprenal.2001.280.5.F860.
- Al Therwani S, Mose FH, Jensen JM, Bech JN, Pedersen EB. Effect of vasopressin antagonism on renal handling of sodium and water and central and brachial blood pressure during inhibition of the nitric oxide system in healthy subjects. BMC Nephrol. 2014;15:100.
- Hager H, Kwon TH, Vinnikova AK, Masilamani S, Brooks HL, Frøkiaer J, et al. Immunocytochemical and immunoelectron microscopic localization of alpha-, beta-, and gamma-ENaC in rat kidney. Am J Physiol Renal Physiol. 2001;280;F1093–106. https://doi.org/10.1152/ajprenal.2001.280.6.F1093.
- Pedersen EB, Eiskjaer H, Madsen B, Danielsen H, Egeblad M, Nielsen CB. Effect of captopril on renal extraction of renin, angiotensin II, atrial natriuretic peptide and vasopressin, and renal vein renin ratio in patients with arterial hypertension and unilateral renal artery disease. Nephrol Dial Transplant. 1993;8:1064–70 http://www.ncbi.nlm.nih.gov/pubmed/8272217. Accessed 8 May 2018.
- Pedersen EB, Danielsen H, Spencer ES. Effect of indapamide on renal plasma flow, glomerular filtration rate and arginine vasopressin in plasma in essential hypertension. Eur J Clin Pharmacol. 1984;26:543–7 http://www. ncbi.nlm.nih.gov/pubmed/6468469. Accessed 8 May 2018.
- Kancir ASP, Pleckaitiene L, Hansen TB, Ekeløf NP, Pedersen EB. Lack of nephrotoxicity by 6% hydroxyethyl starch 130/0.4 during hip arthroplasty. Anesthesiology. 2014;121:948–58. https://doi.org/10.1097/ALN. 000000000000413.
- Song JW, Shim J-K, Kim NY, Jang J, Kwak Y-L. The effect of 0.9% saline versus plasmalyte on coagulation in patients undergoing lumbar spinal surgery; a randomized controlled trial. Int J Surg. 2015;20:128–34. https://doi. org/10.1016/j.ijsu.2015.06.065.
- Young JB, Utter GH, Schermer CR, Galante JM, Phan HH, Yang Y, et al. Saline versus plasma-Lyte a in initial resuscitation of trauma patients: a randomized trial. Ann Surg. 2014;259:255–62. https://doi.org/10.1097/SLA. 0b013e318295feba.

- Kraut JA, Madias NE. Metabolic acidosis: pathophysiology, diagnosis and management. Nat Rev Nephrol. 2010;6:274–85. https://doi.org/10.1038/ nmeph.2010.33.
- Moen V, Brudin L, Rundgren M, Irestedt L. Osmolality and respiratory regulation in humans: respiratory compensation for hyperchloremic metabolic acidosis is absent after infusion of hypertonic saline in healthy volunteers. Anesth Analg. 2014;119:956–64. https://doi.org/10.1213/ANE. 0000000000000404.
- Huang A, Luethi N, Mårtensson J, Bellomo R, Cioccari L. Pharmacodynamics of intravenous frusemide bolus in critically ill patients. Crit Care Resusc. 2017;19:142–9 http://www.ncbi.nlm.nih.gov/pubmed/28651510. Accessed 17 May 2018.
- Hosohata K, Yoshioka D, Tanaka A, Ando H, Fujimura A. Early urinary biomarkers for renal tubular damage in spontaneously hypertensive rats on a high salt intake. Hypertens Res. 2016;39:19–26. https://doi.org/10.1038/hr. 2015.103.
- Gandhi S, Mosleh W, Myers RBH. Hypertonic saline with furosemide for the treatment of acute congestive heart failure: a systematic review and metaanalysis. Int J Cardiol. 2014;173:139–45. https://doi.org/10.1016/j.ijcard.2014.03.020.
- Starklint J, Bech JN, Nyvad O, Jensen P, Pedersen EB. Increased urinary aquaporin-2 excretion in response to furosemide in patients with chronic heart failure. Scand J Clin Lab Invest. 2006;66:55–66. https://doi.org/10.1080/ 00365510500452955.
- Starklint J, Bech JN, Pedersen EB. Urinary excretion of aquaporin-2 after furosemide and felodipine in healthy humans. Scand J Clin Lab Invest. 2005; 65:249–61 http://www.ncbi.nlm.nih.gov/pubmed/16095054. Accessed 28 May 2018.
- Silbert BI, Ho KM, Lipman J, Roberts JA, Corcoran TB, Morgan DJ, et al. Does furosemide increase oxidative stress in acute kidney injury? Antioxid Redox Signal. 2017;26:221–6. https://doi.org/10.1089/ars.2016.6845.
- Bove T, Belletti A, Putzu A, Pappacena S, Denaro G, Landoni G, et al. Intermittent furosemide administration in patients with or at risk for acute kidney injury: meta-analysis of randomized trials. PLoS One. 2018;13: e0196088. https://doi.org/10.1371/journal.pone.0196088.
- Hamishehkar H, Sanaie S, Fattahi V, Mesgari M, Mahmoodpoor A. The effect of furosemide on the level of neutrophil gelatinase-associated lipocalin in critically hospitalized patients with acute kidney injury. Indian J Crit Care Med. 2017;21:442. https://doi.org/10.4103/ijccm.IJCCM_93_17.
- Bagshaw SM, Gibney RTN, Kruger P, Hassan I, McAlister FA, Bellomo R. The effect of low-dose furosemide in critically ill patients with early acute kidney injury: a pilot randomized blinded controlled trial (the SPARK study). J Crit Care. 2017;42:138–46. https://doi.org/10.1016/j.jcrc.2017.07.030.
- 44. Nielsen S, Chou CL, Marples D, Christensen EI, Kishore BK, Knepper MA. Vasopressin increases water permeability of kidney collecting duct by inducing translocation of aquaporin-CD water channels to plasma membrane. Proc Natl Acad Sci U S A. 1995;92:1013–7 http://www.ncbi.nlm. nih.gov/pubmed/7532304. Accessed 28 May 2018.
- Crambert G, Ernandez T, Lamouroux C, Roth I, Dizin E, Martin P-Y, et al. Epithelial sodium channel abundance is decreased by an unfolded protein response induced by hyperosmolality. Physiol Rep. 2014;2:e12169. https:// doi.org/10.14814/phy2.12169.
- Mironova E, Chen Y, Pao AC, Roos KP, Kohan DE, Bugaj V, et al. Activation of ENaC by AVP contributes to the urinary concentrating mechanism and dilution of plasma. Am J Physiol Physiol. 2015;308:F237–43. https://doi.org/ 10.1152/ajprenal.00246.2014.
- Edinger RS, Bertrand CA, Rondandino C, Apodaca GA, Johnson JP, Butterworth MB. The epithelial sodium channel (ENaC) establishes a trafficking vesicle pool responsible for its regulation. PLoS One. 2012;7: e46593. https://doi.org/10.1371/journal.pone.0046593.